

cis-/trans-2-Methoxy-6-phenyl-5,6-dihydro-2H-pyran (14) Mixture. The cis/trans mixture of compound 14 was prepared from benzaldehyde (5.3 g, 50 mmol) and 1-methoxybuta-1,3-diene (8.3 g, 100 mmol) under a pressure of 19.5 kbar at 50 °C in the same way as described for compound 12: yield 6.8 g (80%); bp 119–120 °C (1.2 torr); IR (film) 1660 (C=C), 1160, 1050, 1035 cm⁻¹ (COC); ¹H NMR (CDCl₃) 7.7–7.0 (5 H, m, phenyl), 6.15–5.4 (2 H, m, olefin), 5.12, 4.85 (1 H, 2 br s, cis and trans H-2), 4.66 (1 H, 2 d, H-6), 3.40, 3.30 (3 H, 2 s, cis and trans OCH₃), 2.18 (2 H, m, H-5, H-5'). Anal. Calcd for C₁₂H₁₄O₂: C, 75.7; H, 7.4. Found: C, 75.3; H, 7.4.

6-Phenyl-5,6-dihydro-2-pyrone (4). Compound 4 was obtained from 14 according to the procedure described above for 3: colorless crystals; mp 56–57 °C (hexane); IR (KBr) 1720 (C=O), 1635 (C=C), 1245 cm⁻¹ (COC); ¹H NMR (CDCl₃) 7.6–7.3 (5 H, m, phenyl), 6.99 (1 H, double pd, J_{4,5} + J_{4,5'} = 8.2 Hz, H-4), 6.12 (1 H, dt, J_{3,4} = 8.9, J_{3,5} + J_{3,5'} = 3.5 Hz, H-3), 5.45 (1 H, pd, ΣJ = 16.4 Hz, H-6), 2.62 (2 H, m, H-5, H-5'). Anal. Calcd for C₁₁H₁₀O₂: C, 75.8; H, 5.8. Found: C, 75.8; H, 5.8.

6-Phenyl-2-pyrone (15). A suspension of 360 mg of 10% palladium-on-charcoal in 50 mL of *p*-cymene was dried by azeotropic removal of approximately 5 mL of solvent until the temperature of the vapor was over 170 °C. To this mixture was added 174 mg (1 mmol) of 4, and the resulting suspension was

heated under reflux with stirring for 8 h under a blanket of nitrogen. The mixture was cooled and filtered, and the solvent was removed in vacuo. The residual oily material was purified on a silica gel column with petroleum ether–ethyl acetate (8:2 v/v) to give 79 mg (46%) of the pure α-pyrone 15: mp 65–67 °C (hexane); IR (KBr) 1720 (C=O), 1625 (C=C), 1105, 1070 cm⁻¹ (COC); ¹H NMR (CDCl₃) 8.2–7.1 (5 H, m, phenyl), 7.55 (1 H, invisible, chemical shift was assigned by double-resonance experiment, H-4), 6.71 (1 H, d, J_{3,4} = 7.0 Hz, H-3), 6.33 (1 H, d, J_{4,5} = 9.0 Hz, H-5).

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Registry No. 1a, 25556-21-2; 3, 501-23-5; 4, 2128-90-7; 5, 74986-34-8; 6, 75023-41-5; 7, 74986-35-9; 8, 74986-36-0; 9, 74986-37-1; 10 (isomer 1), 74986-38-2; 10 (isomer 2), 75023-42-6; 11 (isomer 1), 74986-39-3; 11 (isomer 2), 75023-43-7; 12 (trans isomer), 74986-40-6; 12 (cis isomer), 74986-41-7; 13, 74986-42-8; 14 (cis isomer), 74986-41-7; 14 (trans isomer), 74986-40-6; 15, 4660-17-7; methyl undecanoate, 1731-86-8; 1-methoxybuta-1,3-diene, 3036-66-6; *n*-hexanal, 66-25-1; tosylhydrazine, 1709-52-0; benzaldehyde, 100-52-7; *n*-dodecanal, 112-54-9.

Spatane Diterpenoids from the Tropical Marine Alga *Stoechospermum marginatum* (Dictyotaceae)

William H. Gerwick and William Fenical*

Institute of Marine Resources, Scripps Institution of Oceanography, La Jolla, California 92093

M. U. S. Sultanbawa

Department of Chemistry, University of Sri Lanka, Peradeniya Campus, Peradeniya, Sri Lanka

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Ten new metabolites, 2–11, are described as natural products of the tropical marine alga *Stoechospermum marginatum* from Sri Lanka. These new compounds all possess the novel "spatane" tricyclic diterpenoid ring system, and their structures were defined by spectral analyses and by interconversion with derivatives of spatol, a metabolite recently defined fully by X-ray crystallography.

Marine algae of the family Dictyotaceae (Phaeophyta) are prolific producers of interesting secondary metabolites, consisting of C₁₁ acetate-derived compounds,¹ compounds of mixed biosynthesis,^{2–9} sesquiterpenoids,^{10–14} and diterpenoids.^{15–31} The diterpenoids from this group are

particularly unique, as the novel ring systems produced represent unconventional diterpenoid cyclizations not yet

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observed from terrestrial sources. Algae of the family Dictyotaceae are also frequently encountered as the major vegetation in shallow-water tropical and subtropical habitats, even though herbivorous predators are plentiful. Hence, the correlation between secondary metabolite synthesis within this family and predator avoidance seems to be pronounced.

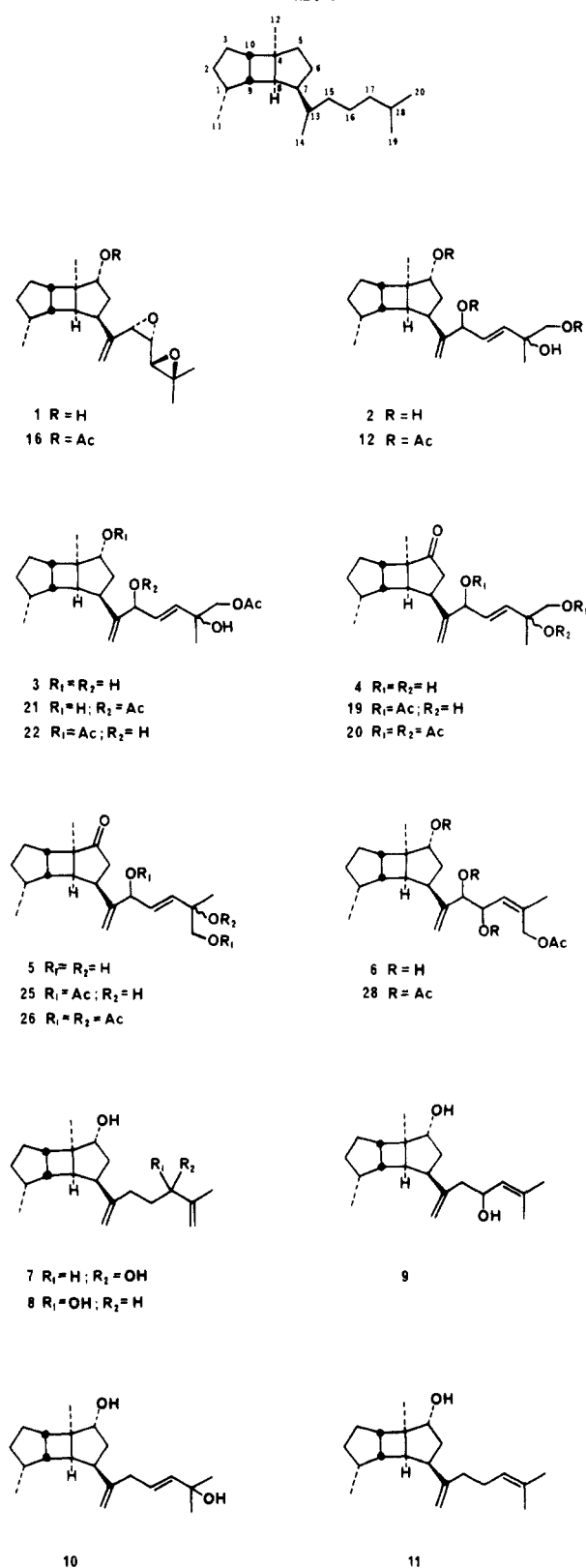
Stoechospermum marginatum (C. Agardh) Kuetzing, an Indian Ocean representative of the family Dictyotaceae, is found in abundance in the warm waters of Sri Lanka. Examination of the organic extracts of *S. marginatum* revealed the alga to contain a complex mixture of new diterpenoids related to the recently described diepoxide spatol (1).³¹ We had earlier isolated spatol from the related alga *Spatoglossum schmittii* from the Galapagos Islands and found this metabolite to possess a tricyclic diterpenoid skeleton related, in part, to the sesquiterpene skeleton of bourbonene.³² Herein, we report the structures of these new spatane diterpenoids as 2-11 (see Chart I). The new compounds have been defined by extensive spectral analysis and by interconversion to degradation products also obtained from spatol.

Conventional silica gel column chromatography of the $\text{CHCl}_3/\text{MeOH}$ extract of air-dried *S. marginatum* provided fractions containing 2-11, mostly as complex mixtures. The most polar fractions, however, contained 2 in nearly pure form, while 3-6 and 7-11 were obtained only after repeated silica gel HPLC.

The most polar and the major metabolite of *S. marginatum* was the tetraol 2, which was isolated as an inseparable mixture of diastereomers ($[\alpha]_D$ (mixture) -10.7°). The tetraol showed strong infrared absorptions at 3500 cm^{-1} , reflecting multiple hydroxyl functionalities, and acetylation ($\text{Ac}_2\text{O}/\text{py}/\text{RT}$) yielded the ester 12 which was confirmed as a triacetate by mass spectrometry and ^1H NMR (Table I). The infrared spectrum of 12 ($\nu_{\text{OH}} = 3500\text{ cm}^{-1}$) showed that a tertiary hydroxyl functionality remained unacetylated. High-resolution mass spectral analysis of 12 failed to illustrate the molecular ion; instead, an $\text{M}^+ - \text{HOAc}$, H_2O fragment was observed. However, the molecular formula for 12, and hence the formula for 2, was readily derived from interpretation of ^{13}C NMR data (Table II). A formula of $\text{C}_{20}\text{H}_{32}\text{O}_4$ (5 unsaturation equivalents) for 2 and $\text{C}_{26}\text{H}_{38}\text{O}_7$ (8 unsaturation equivalents) for the triacetate 12 allowed the deduction that 2 was composed of two double bonds and three carbocyclic rings. Ozonolysis of 12, followed by CH_2N_2 methylation, yielded the fragment 13, which was recognized to possess, intact, the full tricyclic nucleus of compound 2. Compound 13 (Chart II) analyzed for $\text{C}_{16}\text{H}_{24}\text{O}_4$ (5% unsaturation), and possessed secondary acetate and carbomethoxy functionalities. The ^1H and ^{13}C NMR spectra of this degradation product (Tables I and II) indicated that one quaternary and one secondary methyl group were present. Hence, ten carbon atoms remained to construct the tricyclic framework of 13.

The acetate ester function at C-5 in 13 was next selectively saponified with Na_2CO_3 in MeOH to yield the free alcohol 14, which was further utilized for ^1H NMR studies. With incremental additions of $\text{Eu}(\text{fod})_3$ shift reagent (see Experimental Section for details), the protons at various centers in 14 became resolved and, subsequently, inter-related via spin decoupling. Couplings were established and measured between adjacent protons at C-5 through

Chart I



C-10, and protons at C-1 and C-3 were related to C-9 and C-10, respectively. While the methylene pair at C-2 was resolved in this shift study, coupling constants were not reliably measurable. The data thus obtained were highly analogous to the detailed ^1H NMR data obtained from bourbonene,³² therefore indicating an identical *cis-anti-cis* arrangement of the cyclopentane and cyclobutane rings.

For further confirmation that C-5 was a cyclopentane carbon, the alcohol 14 was oxidized with pyridinium

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Table I. Selected ¹H NMR Assignments for *Stoechospermum* Metabolites and Several Related Derivatives^a

| H's at C no. | compound | | | | | | | | | | | | | | | |
|-----------------|---------------------------------|--|-----------------------------|-----------------------------|---|---------------------------------|---------------------------------|---------------------------------|--|---------------------------------|---|---------------------------------|---------------------------------|--|--|--|
| | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 | 16 ^b | | | |
| C ₅ | 3.68 (d, J = 4) | 3.70 (d, J = 4) | | | 3.73 (d, J = 4) | 3.59 (d, J = 4) | 3.73 (d, J = 4) | 3.59 (d, J = 4) | 3.75 (d, J = 4) | 3.73 (d, J = 4) | 4.96 (d, J = 4) | 3.91 (d, J = 4) | 4.85 (d, J = 4) | | | |
| C ₆ | 2.21 (ddd, J = 13, 13, 4) | 2.23 (ddd, J = 13, 13, 4) | 2.95 (dd, J = 13, 13) | 2.95 (dd, J = 13, 13) | 2.23 (ddd, J = 13, 13, 4) | 2.25 (ddd, J = 13, 13, 4) | 2.26 (ddd, J = 13, 13, 4) | 2.15 (ddd, J = 13, 13, 4) | 2.26 (ddd, J = 13, 13, 4) | 2.29 (ddd, J = 13, 13, 4) | 2.32 (ddd, J = 13, 13, 4) | 2.43 (ddd, J = 13, 13, 4) | 2.32 (ddd, J = 13, 13, 4) | | | |
| C ₇ | 2.87 (m) | 2.84 (m) | 2.80 (m) | 2.80 (m) | 2.84 (m) | 2.83 (m) | 2.98 (m) | 2.95 (m) | 2.96 (m) | 2.99 (m) | 2.82 (m) | 3.32 (m) | 2.89 (m) | | | |
| C ₁₁ | 0.86 (d, J = 6) | 0.91 (d, J = 6) | 0.96 (d, J = 6) | 0.96 (d, J = 6) | 0.86 (d, J = 6) | 0.86 (d, J = 6) | 0.84 (d, J = 6) | 0.87 (d, J = 6) | 0.92 (d, J = 6) | 0.86 (d, J = 6) | 0.89 (d, J = 6) | 0.88 (d, J = 6) | 0.95 (d, J = 6) | | | |
| C ₁₂ | 0.93 (s) | 0.99 (s) | 0.96 (s) | 0.96 (s) | 0.98 (s) | 0.98 (s) | 0.98 (s) | 0.91 (s) | 1.00 (s) | 0.98 (s) | 0.88 (s) | 0.95 (s) | 0.90 (s) | | | |
| C ₁₄ | 5.28 (s) | 5.30 (s) | 5.42 (s) | 5.43 (s) | 5.32 (s) | 4.92 (s) | 4.85 (s) | 4.82 (s) | 4.83 (s) | 4.83 (s) | 5.23 (s) | 0.95 (s) | 5.11 (d, J = 1) | | | |
| C ₁₄ | 4.89 (s) | 4.92 (s) | 5.03 (s) | 5.03 (s) | 4.98 (s) | 4.82 (s) | 4.76 (s) | 4.73 (s) | 4.76 (s) | 4.74 (s) | 5.00 (s) | 5.00 (s) | 5.00 (s) | | | |
| C ₁₅ | 4.38 (d, J = 6) | 4.41 (d, J = 6) | 4.52 (d, J = 6) | 4.52 (d, J = 6) | 4.16 (d, J = 4) | 4.05 (dd, J = 6, 6) | 4.04 (dd, J = 6, 6) | | 2.73 (dd, J = 15, 5), 2.59 (dd, J = 15, 5) | | 5.52 (d, J = 6) | 3.30 (d, J = 4) | 3.30 (d, J = 4) | | | |
| C ₁₆ | 5.68 (m) | 5.70 (m) | 5.75 (m) | 5.75 (m) | 4.55 (dd, J = 10, 4) | | | 4.41 (ddd, J = 6, 6, 6) | 5.70 (m) | | 5.80 (dd, J = 16, 6) | | 2.71 (dd, J = 8, 4) | | | |
| C ₁₇ | 5.68 (m) | 5.70 (m) | 5.75 (m) | 5.75 (m) | 5.42 (d, J = 10) | | | 5.09 (d, J = 6) | 5.70 (m) | 5.10 (t, J = 7) | 5.64 (d, J = 16) | | 2.32 (d, J = 8) | | | |
| C ₁₉ | 3.43 (s) | 4.07 (d, J = 10, 3.93 (d, J = 10) | 3.50 (m) | 3.57 (m) | 4.14 (d, J = 13), 4.02 (d, J = 13) | 4.82 (s), 4.74 (s) | 4.76 (s), 4.67 (s) | 1.64 (s) | 1.34 (s) | 1.68 (s) | 4.09 (d, J = 10), 3.98 (d, J = 10) | | 1.41 (s) | | | |
| C ₂₀ | 1.20 (s) | 1.30 (s) | 1.24 (s) | 1.24 (s) | 1.81 (s) | 1.72 (s) | 1.72 (s) | 1.71 (s) | 1.34 (s) | 1.59 (s) | 1.27 (s) | | 1.30 (s) | | | |
| OAc | | 2.09 (s) | | | 2.18 (s) | | | | | | 2.08 (s), 2.05 (s), 2.00 (s) | 1.89 (s) | 1.99 (s) | | | |

^a Spectra were recorded in CDCl₃ at 220 MHz unless otherwise noted, and assignments were aided by spin-decoupling techniques. *J* values are given in hertz and the chemical shifts in δ units. ^b Recorded in CCl₄ solution.

Table II. Tabulated ^{13}C NMR Data for Spatol, *Stoechospermum* Metabolites, and Two Key Derivatives^a

| 1 ^b | 2 ^d | 3 ^c | 4 ^c | 5 ^c | 6 ^c | 7 ^c | 8 ^c | 9 ^b | 10 ^c | 11 ^a | 12 ^b | 13 ^c |
|---------------------|----------------|----------------|----------------|----------------|----------------|----------------|----------------|----------------|-----------------|-----------------|-----------------|-----------------|
| 148.6 s | 150.1 s | 171.7 s | e | 222.4 s | 169.0 s | 148.4 s | 148.5 | 146.2 s | 147.5 s | 148.6 s | 170.3 s | 173.7 s |
| 110.8 t | 136.4 d | 149.6 s | 147.5 s | 148.1 s | 147.1 s | 147.5 s | 147.3 | 133.7 s | 139.8 d | 131.4 s | 169.9 s | 170.4 s |
| | 131.0 d | 135.0 d | 137.3 d | 136.7 d | 139.8 s | 111.0 t | 111.3 | 129.1 d | 124.7 d | 124.4 d | 169.0 s | |
| | 107.2 t | 131.3 d | 131.2 d | 130.8 d | 125.1 d | 108.7 t | 108.7 | 110.8 t | 109.6 t | 108.4 t | 145.5 s | |
| | | 108.8 t | 109.0 t | 109.3 t | 110.2 t | | | | | | 138.6 d | |
| | | | | | | | | | | | 127.4 d | |
| | | | | | | | | | | | 110.2 t | |
| 79.9 d | 78.9 d | 79.8 d | 73.4 s | 73.1 s | 79.9 d | 80.4 d | 80.4 | 80.3 d | 80.3 d | 80.5 d | 81.7 d | 82.1 d |
| 58.4 s | 73.7 d | 73.8 d | 73.1 d | 73.1 d | 75.5 d | 75.7 d | 75.7 | 68.3 d | 70.6 s | | 75.4 d | |
| 58.4 d | 72.2 s | 71.4 s | 69.8 t | 69.7 t | 69.7 d | | | | | | 71.7 s | |
| 57.1 d | 69.5 t | 70.2 t | | | 61.7 t | | | | | | 70.8 t | |
| 54.9 d | | | | | | | | | | | | |
| 47.5 s | 46.5 s | 46.7 s | 48.8 s | 48.9 s | 46.9 s | 46.7 s | 46.8 | 47.0 s | 46.7 s | 46.7 s | 46.6 s | 51.3 q |
| 44.0 d | 43.0 | 43.0 | 42.2 d | 42.3 d | 43.3 | 45.5 d | 45.6 | 46.6 d | 45.4 d | 45.6 d | 43.5 d | 46.6 s |
| 43.6 d | 42.8 | 42.2 | 39.7 | 39.8 | 43.2 | 43.2 d | 43.3 | 44.9 t | 43.2 t | 43.2 d | 43.5 d | 46.4 d |
| 43.5 d ^f | 42.2 | 42.4 | 39.3 | 39.3 | 43.0 | 43.0 d | 43.0 | 43.5 d | 40.8 d | 43.1 d | 37.9 d | 43.9 d |
| 38.0 d | 37.0 | 37.1 | 38.9 | 39.0 | 37.5 | 37.6 t | 37.6 | 43.5 d | 39.1 t | 37.7 d | 37.9 d | 43.0 d |
| 37.2 t ^f | 36.8 | 36.5 | 37.0 | 37.0 | 36.3 | 37.0 | 36.7 | 37.9 d | 37.5 | 36.7 | 36.7 t | 39.8 d |
| 36.8 d | 36.2 | 36.0 | 33.8 t | 33.8 t | 36.3 | 36.7 | 36.5 | 37.1 t | 36.5 | 36.5 | 35.3 d | 36.4 d |
| 35.4 t | 35.0 t | 34.9 t | 27.3 t | 27.3 t | 35.1 | 36.5 t | 35.1 | 36.8 d | 36.5 | 36.4 | 35.3 t | 34.6 t |
| 28.1 t | 27.6 t | 28.8 t | 24.2 q | 24.3 q | 29.7 | 35.1 t | 33.2 | 35.3 t | 35.1 | 35.1 t | 28.0 t | 34.1 t |
| 24.1 q | 23.9 q | 24.2 q | 21.2 q | 21.1 q | 27.9 | 33.2 t | 32.2 | 28.2 t | 29.7 q | 27.9 t | 24.7 q | 27.8 t |
| 19.2 q | 14.0 q | 20.9 q | 14.1 q | 14.0 q | 22.0 q | 32.2 t | 29.7 | 25.7 q | 29.7 q | 26.6 t | 20.8 q | 21.1 q |
| 14.6 q | 12.9 q | 14.1 q | 12.5 q | 12.6 q | 14.3 q | 27.9 t | 27.9 | 18.3 q | 27.9 t | 25.7 q | 20.8 q | 13.5 q |
| 13.4 | | 13.0 q | | | 13.2 q | 17.6 q | 17.5 | 14.6 q | 14.5 q | 17.7 q | 20.5 q | 12.8 q |
| | | | | | | 14.3 q | 14.4 | 13.5 q | 13.1 q | 14.3 q | 14.4 q | |
| | | | | | | 13.1 q | 13.1 | | | 13.1 q | 13.3 q | |

^a Spectra were recorded at 20 MHz in the solvent indicated, and multiplicities were obtained by off-resonance decoupling techniques. ^b Recorded in C_6D_6 solution. ^c Recorded in CDCl_3 solution. ^d Recorded in $(\text{CD}_3)_2\text{CO}$ solution. ^e The carbonyl band was not observed under these instrumental conditions. ^f Assignments may be reversed.

chlorochromate to yield the ketone 15. The ketone showed an infrared carbonyl absorption at 1735 cm^{-1} and a large negative rotation ($[\alpha]_{\text{D}} -226^\circ$) indicative of the five-membered ring with an adjacent asymmetric center at C-4.³³

Unequivocal proof of the structure of the tricyclic nucleus in 2 was obtained by interconversion of both 2 and spatol acetate (16) to the aldehyde 17. Treatment of 16 with H_5IO_6 in ether smoothly cleaved the epoxide, generating the aldehyde 17. Similarly, 2, when treated under analogous conditions, yielded a hydroxy aldehyde, which when acetylated also yielded 17. This latter reaction was apparently facilitated by an allylic rearrangement of the C-18 hydroxyl to C-16. The aldehyde produced from 16 showed $[\alpha]_{\text{D}} +21.3^\circ$, and that from 2 showed $[\alpha]_{\text{D}} +30.1^\circ$, thus indicating that 1 and 2 possess the same absolute stereochemistry at comparable nuclear centers.

The structure of the side chain in 2 followed simply by analysis of ^1H and ^{13}C NMR data. The terminal olefin was positioned at C-13,C-14 on the basis of the results of ozonation and the measurable allylic coupling of the C-7 ring-juncture proton with one proton at C-14. The other disubstituted olefin was assigned as *E* ($J = 15.5\text{ Hz}$) and placed at C-16,C-17 on the basis of the lack of a UV absorption for the metabolite. The remaining spectral features, requiring one tertiary, one secondary, and one primary alcohol, could be assigned only as in 2.

Repeated HPLC failed to separate the two diastereomers which must be epimeric at C-15, or C-18, or at both centers. Cleavage of the C-18,C-19 diol in 2 with NaIO_4 gave the expected methyl ketone 18, which was a single compound ($[\alpha]_{\text{D}} +6.35^\circ$) by HPLC and ^1H NMR analysis. Hence, the epimeric center in 2 must be at C-18, and the structure of 2 is formulated as 5(*R*),15,18(*R* and *S*),19-tetrahydroxyspata-13,16(*E*)-diene.

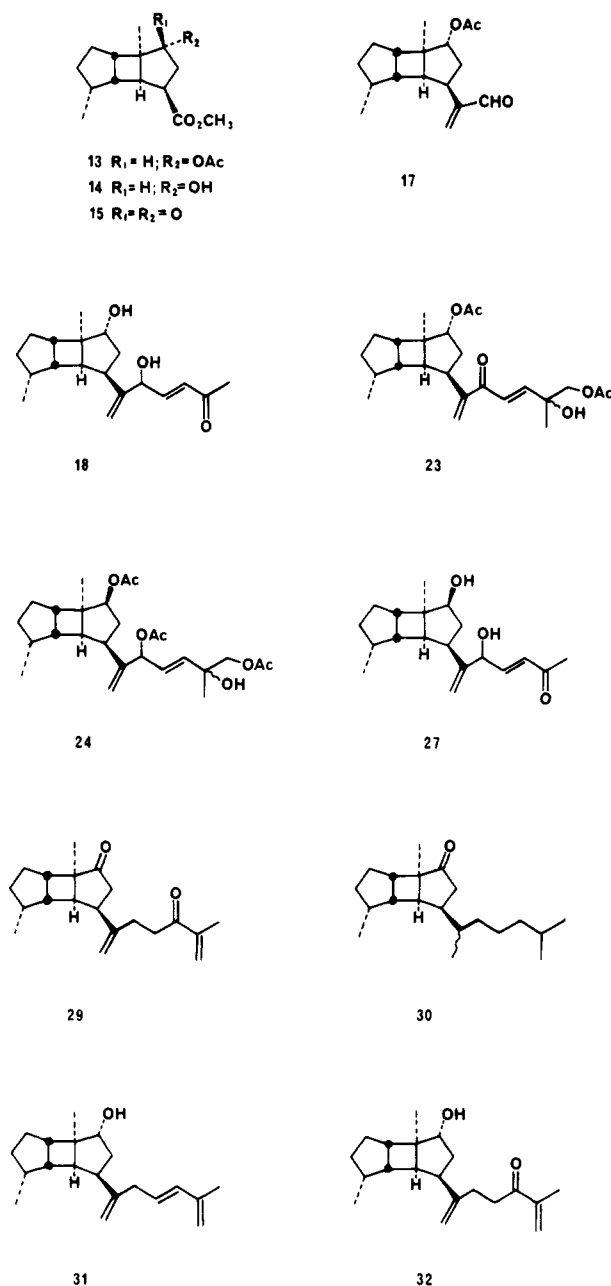
A monoacetate mixture, 3, recognized also to be composed of side-chain diastereomers, was isolated by repeated

HPLC. High-resolution mass spectral analysis showed only an $\text{M}^+ - \text{HOAc} - \text{H}_2\text{O}$ fragment at m/z 300, corresponding to $\text{C}_{20}\text{H}_{26}\text{O}_2$. Consideration of ^1H and ^{13}C NMR data, however, allowed the formulation of 3 as $\text{C}_{22}\text{H}_{34}\text{O}_5$, which is isomeric (minus the acetate) with 2. Acetylation of 3 yielded a triacetate, 12, which was identical, including optical rotation, with that produced from 2. The position of acetylation in 3 could readily be deduced to be at C-19 by ^1H NMR, since the primary acetate AB pattern was observed at δ 4.07 (d, $J = 10\text{ Hz}$) and 3.93 (d, $J = 10\text{ Hz}$) (Table I), as was the case for the triacetate 12. Therefore, 3 is assigned in analogy with 2 as the identical diastereomeric mixture at C-18.

Two epimeric ketones, 4 and 5, were isolated from the extract of *S. marginatum*, and in this case the mixture of diastereomers was successfully separated by silica gel HPLC. Ketone 4, an oil, showed $[\alpha]_{\text{D}} -136.5^\circ$ and analyzed for $\text{C}_{20}\text{H}_{30}\text{O}_4$ by combined ^{13}C NMR and high-resolution mass spectral methods. The ketone was readily recognized as being cyclopentanoid, and therefore placed at C-5, by its characteristic infrared absorption at 1730 cm^{-1} . This assignment was reinforced by ^{13}C NMR data, since the strained carbonyl carbon was observed at 222.4 ppm. Interpretation of additional ^1H and ^{13}C NMR data indicated the identical side-chain substitution in 4 as found in 2 and 3. Acetylation of 4 yielded a 4:1 mixture of the diacetate 19 and the triacetate 20. The diacetate thus obtained formed the basis of a precise comparison with 3. Selective acetylation of 3 ($\text{Ac}_2\text{O}/\text{py}/0^\circ\text{C}/3\text{ h}$) gave a mixture of diacetates 21 and 22, which were further oxidized with pyridinium chlorochromate to yield the keto diacetates 19 and 23. The keto acetate 19 was separated by recycle-HPLC and found to be identical with the acetate produced from 4. During the HPLC separation of 19 and 23, the diastereomers at C-18 were rendered separable. We are not able, however, to assess which epimer was isolated, nor are we able to establish the absolute stereochemistry at C-18 in 4. The ketone 4 was therefore assigned as 5-oxo-15,18(*R**),19-trihydroxyspata-13,16(*E*)-diene.³⁴

(33) Crabbé, P. "Optical Rotary Dispersion and Circular Dichroism in Organic Chemistry"; Holden-Day: San Francisco, 1965; p 378.

Chart II



Reduction of the ketone 4 with $Li(t\text{-}BuO)_3AlH$ in ethanol, followed by acetylation, yielded the triacetate 24. This reduction selectively produced the nonnatural C-5 β -alcohol, as could be deduced from the C-5 methine proton NMR characteristics. In 24, the C-5 proton is observed as a doublet of doublets with couplings of 8 and 10 Hz. In contrast, the epimeric C-5 proton in the natural products 1-3 and 6-11 is limited to a single 4-Hz coupling with the C-6 α -hydrogen. This observation provided considerable support in subsequent stereochemical assignments of derivatives at C-5.

The epimeric ketone 5 showed spectral characteristics (1H and ^{13}C NMR, mass, IR) nearly superimposable with those of 4, except for exhibiting an optical rotation of $[\alpha]_D -99.2^\circ$. As in the case of 2, the question of epimerization

at C-15 or C-18 remained. Acetylation of 5 produced the diacetate 25 and the triacetate 26 in a 4:1 ratio. The 1H NMR features of 25 were nearly identical with those of 19, reaffirming the epimeric relationship of 4 and 5. To establish the site of epimerization, 5 was reduced with $LiAlH_4$, and the crude tetraol mixture was treated with $NaIO_4$ to yield the methyl ketones 18 and 27. The methyl ketone 18 could be readily separated and was found to be identical ($[\alpha]_D +10.1^\circ$) with that produced from 2. Ketones 4 and 5 are, therefore, shown to be epimeric at C-18, and via the favorable comparison with 2, 5 may be described as 5-oxo-15,18(S^*),19-trihydroxyspata-13,16(E)-diene.³⁴

A polar monoacetate, 6, recognized to be isomeric with 3, was isolated by repeated HPLC of the more polar fractions. As with 3, high-resolution mass spectrometry showed only $M^+ - HOAc - H_2O$ fragments, and the molecular composition was fully established as $C_{22}H_{34}O_5$ by ^{13}C NMR. As the tricyclic nucleus of 6 appeared to be identical with those of 1-5 by spectral analysis, the isomeric relationship was, here also, assigned within the side chain. Cleavage ($NaIO_4$) of the side-chain vicinal diol at C-15,C-16 yielded an aldehyde product, which when acetylated was identical with 17 ($[\alpha]_D +26.5^\circ$) as produced from both 1 and 2. The composition of the tricyclic nucleus was, therefore, firmly established. 1H NMR analysis (Table I) showed 6 to possess one olefinic methyl group (δ 1.81, 3 H, s) and one trisubstituted olefin (δ 5.42, 1 H, d). The coupling of the olefin proton with the alcohol methine proton at C-16 ($J = 10$ Hz) further established the diol at C-15,C-16 and allowed the formulation of the side-chain structure. The single acetate was clearly at C-19, since an isolated AB pattern at δ 4.14 and 4.02 ($J = 13$ Hz) was characteristic of the shift expected for a primary acetate. The stereochemistry of the C-17,C-18 olefin was tentatively assigned on the basis of a comparison of the ^{13}C NMR calculated values for E and Z geometrical isomers with that measured for 6.³⁵ A calculated value of 21.9 ppm for the Z isomer is in rough agreement with our measured shift of >22 ppm, thus supporting the Z assignment.

The epimeric alcohols 7 and 8 were separable by HPLC and were isolated from the medium-polarity column chromatography fractions. Compound 7, an optically active oil ($[\alpha]_D +35.2^\circ$), analyzed as a diol of molecular formula $C_{20}H_{32}O_2$ by combined spectral methods. The 5 unsaturation equivalents inherent in this formula could be accounted for by two olefinic bonds (both terminal) and the familiar tricyclic skeleton. An interconversion with degradation products from 1 could not be accomplished with 7; however, complete analysis of 1H and ^{13}C NMR data gave very convincing evidence in support of structure 7. In the 1H NMR spectrum, the characteristic methine protons of the spatane nucleus were clearly visible and possessed the same coupling constants. The superimposability of all comparable ^{13}C NMR bands further supported this assignment. What remained was to establish the structure of the side chain, and this was accomplished by 1H NMR decoupling experiments and by chemical modification. Two terminal olefinic bonds were readily established by ^{13}C NMR, and the presence of an olefinic methyl group in the 1H NMR spectrum secured the placement of olefins at C-13,C-14 and C-18,C-19. What

(34) The R^*,S^* notation is used here as an aid in nomenclature to indicate the site of epimerization for two diastereomeric molecules. In 4 and 5 the notation indicates a C-18 epimeric relationship, while in 7 and 8 the epimeric relationship lies at C-17. A more suitable method for naming compounds in this circumstance does not appear to exist.

(35) Calculated ^{13}C NMR shifts for the C-20 methyl group with the C-17, C-18 E and Z olefin involved the model compounds geraniol, the natural product 9, and 2,6-dimethyl-1-hydroxyocta-2(E),7-diene. The E isomer yielded a value of 14.6 ppm while the Z isomer calculated for 21.9 ppm; see: Wehrli, F. W.; Nishida, T. in "Progress in the Chemistry of Organic Natural Products"; Herz, W., Grisebach, H., Kirby, G. W., Eds.; Wien, Springer-Verlag: New York, 1979; p 24.

remained was to establish the position of the side-chain alcohol at one of three possible carbon sites. Pyridinium chlorochromate oxidation of **7** yielded the α,β -unsaturated ketone derivative **29**, which was readily assignable on the basis of the shifts observed in the terminal olefin protons at C-19. As expected for the deshielded β -carbon of the enone, the C-19 protons were shifted from δ 4.82 and 4.74 in **7** to δ 6.05 and 5.84 in **29**. Further, the C-20 methyl group was, expectedly, also shifted to δ 1.94. The diketone **29** also exhibited a large negative rotation ($[\alpha]_D -190^\circ$) in close accord with the rotation found for the chiral C-5 ketone in **15**. On the basis of these chemical and spectral analyses, the structure for **7** is formulated as 5(*R*),17-(*R*^{*})-dihydroxyspata-13,18-diene.³⁴

The epimeric diol **8** showed $[\alpha]_D +18.4^\circ$ and was virtually superimposable spectrally with diol **7**. Since the ¹H NMR features of the C-5 alcohol methine proton were so clearly in support of the 5 α -hydroxyl, the epimeric relationship of **8** was envisioned to be at C-17. Confirmation of the epimeric relationship at this center was obtained via oxidation of **8** to yield **29**. The diketone obtained was identical in all respects ($[\alpha]_D -185^\circ$) with that from **7**, thus establishing **8** as 5(*R*),17(*S*^{*})-dihydroxyspata-13,18-diene.³⁴

An isomeric diol, **9**, was also purified from the medium-polarity column fractions. Compound **9**, an oil, showed $[\alpha]_D +6.3^\circ$ and analyzed for C₂₀H₃₂O₂ by combined ¹³C NMR and high-resolution mass spectral methods. The ¹H and ¹³C NMR features of **9** again clearly delineated the tricyclic nucleus, and spectral features to assign the elements of the side chain were also apparent. In addition to the consistent *exo*-methylene at C-13,C-14, the ¹H NMR spectrum showed two olefinic methyls at δ 1.71 and 1.64, a single olefin proton at δ 5.09, and an allylic alcohol methine proton at δ 4.41. This latter band was found to be coupled to the olefin proton at δ 5.09 ($J = 6$ Hz); hence, the allylic alcohol and olefin constellation were securely placed at the terminal position in the side chain.

Ozonolysis of **9**, followed by CH₂N₂ methylation and acetylation (Ac₂O/py), yielded **13** unexpectedly, in only modest yield. It would appear that dehydration occurred prior to ozonolysis or that more complex mechanisms afforded oxidative cleavage of the C-13,C-15 bond. The ester produced from **9** showed $[\alpha]_D +1.75^\circ$, thus indicating the nuclear stereochemistry of **9** to be identical with that of **1** and **2**. In addition, selective dehydration of the allylic alcohol in **9**, followed by hydrogenation and pyridinium chlorochromate oxidation, produced the saturated ketone **30**. The ketone showed a large negative rotation ($[\alpha]_D -101.2^\circ$), consistent with **15** and **29**. The diol **9** can therefore be assigned as 5(*R*),16-dihydroxyspata-13,17-diene.

HPLC separation of another medium-polarity column fraction yielded an additional isomeric diol assigned as **10**. The diol showed $[\alpha]_D +1.9^\circ$, and here also combined spectral methods indicated a molecular formula of C₂₀H₃₂O₂. Analysis of spectral data in a consistent fashion suggested the tricyclic nucleus to be present as well as indicated a side-chain composition isomeric with **9**. Infrared absorptions at 975 cm⁻¹ and a two-proton olefin band at δ 5.70 suggested a trans-disubstituted double bond to be present. Also, two methyl groups were observed in the ¹H NMR spectrum at δ 1.34, suggesting the placement of these methyls at a carbon bearing the remaining hydroxyl group. Since the diol **10** lacked a UV chromophore, the olefin was securely placed at C-16,C-17, resulting in the final assignment of diol **10**. To confirm the relationship of **10** to **9**, and hence of diol **10** to both **1** and **2**, the compound was dehydrated with *p*-toluenesulfonic acid in

aqueous acetone to yield the triene **31**. Hydrogenation of the triene followed by pyridinium chlorochromate oxidation gave the ketone **30** ($[\alpha]_D -99.5^\circ$) which was identical with the compound produced from **9**. Since **9** had been rigorously related to **1** and **2**, diol **10** may be formulated as 5(*R*),18-dihydroxyspata-13,16(*E*)-diene.

Evaluation of the relatively nonpolar column fractions from *S. marginatum* yielded only one monoalcohol, **11**. Alcohol **11** showed $[\alpha]_D +29.4^\circ$ and analyzed for C₂₀H₃₂O by ¹³C NMR and high-resolution mass spectrometry. As with all the compounds from this source, the tricyclic portion of the molecule seemed intact by evaluation of ¹H and ¹³C NMR data. In addition to these spectral features, several ¹³C and ¹H NMR bands confirmed the presence of an additional trisubstituted double bond. Further, ¹³C NMR bands at 124.2 (d) and 131.4 (s) ppm and ¹H NMR singlet resonances (3 H each) at δ 1.68 and δ 1.59 indicated the olefin was substituted with two methyl groups. In the ¹H NMR spectrum the single olefin proton appeared at δ 5.10 as a characteristic triplet ($J = 7$ Hz), indicating an adjacent methylene group. From these data the olefin was confidently placed at C-17,C-18 to complete the structure proposal for **11**.

Treatment of **11** with photochemically produced singlet oxygen (methylene blue sensitizer) produced the unsaturated ketone **32**, in modest yield, after the expected "ene" reaction and apparent autocatalytic decomposition of the intermediate hydroperoxide at C-17. Ketone **32** was further oxidized with pyridinium chlorochromate to yield the diketone **29**. The diketone showed $[\alpha]_D -150^\circ$ and was identical with that produced from **7** and **8**.

The alcohol **11** was also converted to the ketone **30** as the final confirmation of structure. Hydrogenation (Pt/Et₂O), followed by pyridinium chlorochromate oxidation gave the ketone **30** ($[\alpha]_D -110^\circ$), identical in all respects with that produced from **9** and **10**. The structure of alcohol **11** is, therefore, securely assigned as 5(*R*)-hydroxyspata-13,17-diene.³⁶

Experimental Section

Infrared spectra were recorded on a Perkin-Elmer Model 137 spectrophotometer, and optical rotations were measured on a Perkin-Elmer Model 141 polarimeter using a 10-cm microcell. ¹H NMR spectra were recorded on Varian HR-220 and T-60 spectrometers and on a home-built 360-MHz spectrometer, and ¹³C NMR spectra were recorded on a Varian CFT-20 spectrometer. All chemical shifts are reported relative to Me₄Si (δ 0), and coupling constants are in hertz. Low-resolution mass spectra (LRMS) were obtained on a Hewlett-Packard 5930-A mass spectrometer, and high-resolution mass spectra (HRMS) were obtained through the Department of Chemistry, University of California, Los Angeles. All solvents used were distilled from glass prior to use.

Collection, Extraction, and Chromatography. *Stoecho-spermum marginatum* was collected in shallow water along the coast of Sri Lanka in 1977 and air-dried prior to extraction (3.2 kg). A cold CHCl₃/MeOH (2/1) extraction yielded 120 g of extract after solvent evaporation, and a subsequent extraction with warm solvent (~50 °C) yielded an additional 82 g. No significant differences were observed between the two extracts.

Conventional silica gel column chromatography using various proportions of CH₂Cl₂, EtOAc, and MeOH gave fractions rich in 2-11. The least polar fractions (2% EtOAc/CH₂Cl₂) contained **11**, while moderate polarity fractions (20% EtOAc/CH₂Cl₂) contained 7-10. Compound **3** was eluted with 100% EtOAc. The more polar compounds were eluted with 25% MeOH/EtOAc and

(36) We have recently determined, by direct comparison, that compound **11** is identical with the previously reported diterpenoid stoecho-spermol, also isolated from *S. marginatum*.¹⁹ Since the structure of diterpenoid **11** has been confidently assigned, the structure of stoecho-spermol should be accordingly revised.

50% MeOH/EtOAc and consisted of 4–6 (25%) and 2 (50%). Each compound was subsequently purified by high-performance liquid chromatography (HPLC) on a 16 mm × 50 cm preparative silica gel column by using various proportions of EtOAc in iso-octane.

Acetylations. Conversions of the natural products and several degradation products into their corresponding acetate esters were accomplished in the following general way. Excess acetic anhydride and dry pyridine were added to milligram quantities of natural products, and the resultant solution was stirred at room temperature overnight. The reaction was quenched with ice and water, and the reaction products were extracted with Et₂O (3×). The ether phase was washed with 5% HCl (3×), water (3×), and saturated NaHCO₃ (3×), dried over anhydrous MgSO₄, and reduced in vacuo to yield the crude esters. Purification, when necessary, was provided by HPLC under the conditions mentioned above.

5(R),15,18(R and S),19-Tetrahydroxypata-13,16(E)-diene (2). The tetraol mixture was isolated as 5.3% of the extract. The mixture was an oil with the following spectral features: $[\alpha]_D -10.7^\circ$ (c 1.82, CHCl₃); IR (CHCl₃) 3450, 2950, 1710, 1450, 1370, 1250, 1025, 940, 908 cm⁻¹. Acetate 12 was purified by using HPLC (50% EtOAc/iso-octane), as an oil: $[\alpha]_D -36^\circ$ (c 1.32, CHCl₃); IR (CHCl₃) 3550, 2950, 1730, 1650, 1460, 1375, 1250, 1040, 970, 910 cm⁻¹; LRMS (70 eV, 200 °C), *m/z* (relative intensity) 402 (0.2), 384 (0.3), 342 (0.3), 269 (1.0), 260 (23), 225 (19), 200 (16), 187 (40), 135 (79), 81 (100); HRMS (70 eV, 180 °C), *m/z* (relative intensity) 384.2295 (M⁺ - HOAc - H₂O, C₂₄H₂₂O₄, 5.0, -0.6 mamu dev), 342.2195 (C₂₀H₃₀O₃, 9.3), 269.1905 (C₁₉H₂₅O, 42), 260.1417 (C₁₆H₂₀O₃, 90), 225.1638 (C₁₇H₂₁, 36), 200.1206 (C₁₄H₁₆O, 38), 187.1112 (C₁₃H₁₅O, 56), 145.1024 (C₁₁H₁₃, 54), 143.0866 (C₁₁H₁₁, 56), 135.1171 (C₁₀H₁₅, 100).

19-Acetoxy-5(R),15,18(R and S)-trihydroxypata-13,16(E)-diene (3). The monoacetate diastereomeric mixture was isolated as an oil as 1.4% of the extract and showed the following spectral features: $[\alpha]_D -16.1^\circ$ (c 1.48, CHCl₃); IR (CHCl₃) 3500, 2975, 1735, 1650, 1450, 1375, 1240, 1110, 1040, 940, 908 cm⁻¹; HRMS (70 eV, 180 °C), *m/z* (relative intensity) 300.2080 (M⁺ - HOAc - H₂O, C₂₀H₂₆O₂, -1.0 mamu dev, 6), 287.1983 (C₁₉H₂₇O₂, 13), 269.1888 (C₁₉H₂₅O, 14), 211.1474 (C₁₆H₁₉, 14), 159.1182 (C₁₂H₁₅, 27), 145.1007 (C₁₁H₁₃, 35), 135.1175 (C₁₀H₁₅, 68), 105.0711 (C₈H₉, 54), 81.0319 (C₅H₅O, 100). Acetate 12. Standard acetylation conditions of 7.4 mg of 3 (0.0196 mmol) followed by preparative thin-layer chromatography yielded 5.4 mg (0.0117 mmol, 60%) of 12, which showed $[\alpha]_D -41.3^\circ$ (c 0.54, CHCl₃) and which was identical with 12 produced from 2 by 220-MHz ¹H NMR, IR, and LRMS.

5-Oxo-15,18(R or S),19-trihydroxypata-13,16(E)-diene (4). The ketone was isolated as an oil by HPLC (100% EtOAc) as 0.23% of the extract. Ketone 4 showed the following spectral characteristics: $[\alpha]_D -136.5^\circ$ (c 1.22, CHCl₃); IR (CHCl₃) 3450, 2950, 1730, 1450, 1380, 1240, 1110, 1040, 970, 910 cm⁻¹; HRMS (70 eV, 180 °C), *m/z* (relative intensity) 303.1956 (M⁺ - CH₃O, C₁₉H₂₇O₃, -0.4 mamu dev, 66), 298.1947 (C₂₀H₂₆O₂, 2), 285.1852 (C₁₉H₂₅O₂, 51), 227.1454 (C₁₆H₁₉O, 41), 199.1498 (C₁₅H₁₉, 22), 177.1269 (C₁₂H₁₇O, 24), 153.0928 (C₉H₁₃O₂, 73), 151.1132 (C₁₀H₁₅O, 100), 149.0962 (C₁₀H₁₃O, 43), 99.0448 (C₅H₇O₂, 52), 81.0691 (C₆H₉, 98). Acetates 19 and 20. Standard acetylation of 15.0 mg (0.045 mmol) of 4 followed by preparative thin-layer chromatography provided in a ratio of 4:1 the desired diacetate 19 (15.0 mg, 0.358 mmol, 80%) and unexpected triacetate 20 (1.96 mg, 0.009 mmol, 20%). Compound 19 produced from 4 showed the following: $[\alpha]_D -104.0^\circ$ (c 1.50, CHCl₃); ¹H NMR (220 MHz, CDCl₃) δ 5.85 (1 H, d, *J* = 13), 5.66 (2 H, m), 5.37 (1 H, s), 5.08 (1 H, s), 4.11 (1 H, d, *J* = 10), 3.96 (1 H, d, *J* = 10), 2.87 (2 H, m), 2.76 (1 H, m), 1.4–2.5 (8 H, m), 2.08 (3 H, s), 2.07 (3 H, s), 1.33 (3 H, s), 0.99 (3 H, s), 0.92 (3 H, d, *J* = 7). The triacetate 20 displayed the following ¹H NMR bands (220 MHz, CDCl₃): 5.96 (1 H, d, *J* = 13), 5.60 (2 H, m), 5.37 (1 H, s), 5.08 (1 H, s), 4.26 (2 H, s), 2.91 (2 H, m), 2.80 (1 H, m), 1.2–2.5 (8 H, m), 2.11 (3 H, s), 2.07 (3 H, s), 2.01 (3 H, s), 1.57 (3 H, s), 0.96 (3 H, s), 0.93 (3 H, d, *J* = 7).

5-Oxo-15,18(S or R),19-trihydroxypata-13,16(E)-diene (5). Ketone 5 was isolated along with 4 by HPLC. The ketone composed 0.4% of the extract and showed the following spectral features: $[\alpha]_D -99.2^\circ$ (c 1.84, CHCl₃); IR (CHCl₃) 3450, 2975, 1730, 1460, 1375, 1240, 1030, 965, 910 cm⁻¹; HRMS (70 eV, 180 °C), *m/z*

(relative intensity) 303.1986 (obsd M⁺ - CH₃O, C₁₉H₂₇O₃, 2.6 mamu dev, 1.4), 285.1845 (C₁₉H₂₅O₂, 1.5), 227.1415 (C₁₆H₁₉O, 7), 161.0965 (C₁₁H₁₃O, 15), 159.1163 (C₁₂H₁₅, 18), 153.0913 (C₉H₁₃O₂, 34), 151.1132 (C₁₀H₁₅O, 51), 149.0974 (C₁₀H₁₃O, 27), 99.0427 (C₅H₇O₂, 46), 93.0709 (C₇H₉, 43), 81.0702 (C₈H₉, 100). Acetates 25 and 26. Standard acetylation of 16.2 mg (0.0485 mmol) of 5, followed by preparative thin-layer chromatography, provided two compounds, 25 (14.4 mg, 0.0344 mmol, 71%) and 26 (3.9 mg, 0.0086 mmol, 17.7%). The diacetate 25 showed the following: $[\alpha]_D -79.0^\circ$ (c 1.44, CHCl₃); ¹H NMR (220 MHz, CDCl₃) δ 5.81 (1 H, d, *J* = 14), 5.65 (2 H, m), 5.36 (1 H, s), 5.05 (1 H, s), 4.09 (1 H, d, *J* = 10), 3.94 (1 H, d, *J* = 10), 2.88 (2 H, m), 2.77 (1 H, m), 1.4–2.5 (8 H, m), 2.09 (3 H, s), 2.06 (3 H, s), 1.30 (3 H, s), 0.95 (3 H, s), 0.89 (3 H, d, *J* = 7). The triacetate 26 displayed the following ¹H NMR bands (220 MHz, CDCl₃): δ 5.95 (1 H, d, *J* = 15), 5.61 (1 H, d, *J* = 8), 5.50 (1 H, dd, *J* = 15, 8), 5.36 (1 H, s), 5.05 (1 H, s), 4.23 (2 H, s), 2.88 (2 H, m), 2.77 (1 H, m), 1.5–2.4 (8 H, m), 2.10 (3 H, s), 2.08 (3 H, s), 2.00 (3 H, s), 1.55 (3 H, s), 0.95 (3 H, s), 0.89 (3 H, d, *J* = 7).

19-Acetoxy-5(R),15,16-trihydroxypata-13,17(Z)-diene (6). The monoacetate 6 was isolated as an oil (0.26% extract) by HPLC (100% EtOAc) and showed the following spectral features: $[\alpha]_D -8.7^\circ$ (c 1.22, CHCl₃); IR (CHCl₃) 3450, 2950, 1720, 1640, 1440, 1370, 1240, 1040, 1000, 940, 908 cm⁻¹; HRMS (70 eV, 180 °C), *m/z* (relative intensity) 301.2158 (obsd M⁺ - OAc - H₂O, C₂₀H₂₆O₂, -1.0 mamu dev, 1.3), 136.0362 (C₉H₁₂O, 100), 135.1157 (C₁₀H₁₅, 69), 107.0866 (C₈H₁₁, 55), 101.0596 (C₈H₉O₂, 69), 93.0703 (C₇H₉, 60), 81.0321 (C₅H₅O, 80). Acetate 28. Standard acetylation of 12 mg (0.032 mmol) of 6 followed by preparative thin-layer chromatography gave pure 28, which displayed the following absorptions in the ¹H NMR (220 MHz, CDCl₃): δ 5.80 (1 H, dd, *J* = 10, 5), 5.51 (1 H, d, *J* = 5), 5.45 (1 H, d, *J* = 10), 5.23 (1 H, s), 5.02 (1 H, s), 4.93 (1 H, d, *J* = 4), 4.66 (1 H, d, *J* = 12), 4.54 (1 H, d, *J* = 12), 2.86 (1 H, m), 2.43 (1 H, ddd, *J* = 13, 13, 4), 1.1–2.2 (9 H, m), 2.25 (3 H, s), 2.20 (3 H, s), 2.19 (3 H, s), 2.17 (3 H, s), 1.93 (3 H, s), 0.95 (3 H, s), 0.83 (3 H, d, *J* = 6).

5(R),17(S*)-Dihydroxypata-13,18-diene (7). The diol 7, an oil, was isolated by HPLC (50% EtOAc/iso-octane) as 0.22% of the extract and showed the following spectral features: $[\alpha]_D +35.2^\circ$ (c 0.83, CHCl₃); IR (CHCl₃) 3500, 2975, 1725, 1440, 1380, 1240, 1100, 910, 840 cm⁻¹; HRMS (70 eV, 180 °C), *m/z* (relative intensity) 271.2053 (obsd M⁺ - CH₃O (H₂O, CH₃), C₁₉H₂₇O, -0.9 mamu dev, 6.2), 204.1491 (C₁₄H₂₀O, 74), 187.1470 (C₁₄H₁₈, 33), 159.1189 (C₁₂H₁₅, 35), 151.1110 (C₁₀H₁₅O, 27), 133.1022 (C₁₀H₁₃, 39), 120.0940 (C₉H₁₂, 77), 93.0699 (C₇H₉, 80), 81.0702 (C₆H₉, 100), 79.0551 (C₆H₇, 60).

5(R),17(R*)-Dihydroxypata-13,18-diene (8). Diol 8, an oil, was isolated by HPLC (50% EtOAc/iso-octane) as 0.38% of the extract and showed the following spectral characteristics: $[\alpha]_D +18.4^\circ$ (c 0.67, CHCl₃); IR (CHCl₃) 3500, 3590, 1725, 1640, 1440, 1370, 1230, 1110, 1030, 985, 940, 905 cm⁻¹; HRMS (70 eV, 180 °C), *m/z* (relative intensity) 205.1563 (obsd M⁺ - C₆H₁₁O (CH₂C-H₂CH(OH)C(CH₂)CH₃), C₁₄H₂₁O, -2.9 mamu dev, 20), 204.1514 (C₁₄H₂₀O, 66), 161.1326 (C₁₂H₁₇, 30), 120.0942 (C₉H₁₂, 82), 107.0863 (C₈H₁₁, 73), 105.0706 (C₈H₉, 61), 93.0709 (C₇H₉, 75), 81.0692 (C₆H₉, 100), 79.0537 (C₆H₇, 75).

5(R),16-Dihydroxypata-13,17-diene (9). The diol 9, an oil, was isolated by HPLC (50% EtOAc/iso-octane) as 0.05% of the extract and showed the following spectral features: $[\alpha]_D +6.3^\circ$ (c 1.91, CHCl₃); IR (CCl₄) 3350, 3050, 1650, 1450, 1380, 1185, 1115, 1050, 945, 900 cm⁻¹; LRMS (75 °C, 35.5 eV), *m/z* (relative intensity) 304 (M⁺, 0.8), 286 (1.5), 202 (5.3), 135 (37), 86 (100); HRMS (70 eV, 180 °C), *m/z* (relative intensity) 286.2300 (obsd M⁺ - H₂O, C₂₀H₃₀O, 0.3 mamu dev, 2.6), 204.1519 (C₁₄H₂₀O, 27), 135.1174 (C₁₀H₁₅, 60), 107.0878 (C₈H₁₁, 41), 105.0707 (C₈H₉, 45), 93.0696 (C₇H₉, 47), 91.0537 (C₇H₇, 49), 85.0651 (C₅H₅O, 100), 81.0704 (C₆H₉, 69).

5(R)-18-Dihydroxypata-13,16(E)-diene (10). The diol 10, an oil, was isolated (0.3% extract) by HPLC (50% EtOAc/iso-octane) and showed the following spectral features: $[\alpha]_D +1.9^\circ$ (c 1.35, CHCl₃); IR (CHCl₃) 3500, 3000, 1650, 1460, 1370, 1140, 1110, 1035, 975, 940, 895 cm⁻¹; HRMS (70 eV, 180 °C), *m/z* (relative intensity) 286.2309 (obsd M⁺ - H₂O, C₂₀H₃₀O, 1.2 mamu dev, 3.3), 268.2195 (C₂₀H₂₈, 20), 204.1515 (C₁₄H₂₀O, 93), 145.1005 (C₁₁H₁₃, 55), 119.0866 (C₈H₁₁, 76), 105.0699 (C₈H₉, 100), 91.0553 (C₇H₇, 88), 81.0711 (C₆H₉, 92).

5(R)-Hydroxypata-13,17-diene (11). The alcohol 11, an oil (1.3% of the extract), was isolated in pure form in the column chromatography process and showed the following spectral features: $[\alpha]_D^{25} +29.4^\circ$ (c 1.07, CHCl_3); IR (CHCl_3) 3500, 2950, 1650, 1440, 1370, 1110, 1040, 985, 940, 895 cm^{-1} ; LRMS (35.5 eV), m/z (relative intensity) 288 (obsd, M^+ , 0.8), 270 (0.8), 259 (1.2), 227 (2.0), 206 (23), 145 (18), 137 (30), 121 (22), 107 (30), 81 (50), 69 (100), 42 (67); HRMS (70 eV, 180 $^\circ\text{C}$), m/z (relative intensity) 270.2341 (obsd $\text{M}^+ - \text{H}_2\text{O}$, $\text{C}_{20}\text{H}_{30}$, -0.7 mamu dev, 61), 206.1679 ($\text{C}_{14}\text{H}_{22}\text{O}$, 65), 147.1168 ($\text{C}_{11}\text{H}_{15}$, 30), 137.0971 ($\text{C}_9\text{H}_{13}\text{O}$, 87), 135.1179 ($\text{C}_{10}\text{H}_{15}$, 70), 105.0710 (C_8H_9 , 53), 93.0700 (C_7H_9 , 56), 91.0541 (C_7H_7 , 58), 81.069 (C_6H_9 , 100).

H_2IO_6 Degradation of 16 To Yield Aldehyde 17. A 5-mL sample of a saturated ether solution of H_2IO_6 was added dropwise to 54.9 mg (0.152 mmol) of 16 in 3 mL of ether over 0.5 h at 0 $^\circ\text{C}$. The solution was allowed to warm to room temperature and further stirred for an additional 48 h. Filtration and removal of ether in vacuo yielded a complex oily mixture which was separated by HPLC (10% EtOAc/isooctane, μ -Porasil column). The major product produced was 17: 4.8 mg (0.014 mmol, 11.5%); $[\alpha]_D^{25} +21.3^\circ$ (c 0.48, CHCl_3); $^1\text{H NMR}$ (220 MHz, CDCl_3) δ 9.50 (1 H, s), 6.22 (1 H, s), 6.09 (1 H, s), 4.95 (1 H, d, $J = 4.3$), 3.36 (1 H, dddd, $J = 5, 5, 13, <1$), 2.39 (1 H, ddd, $J = 13, 13, 4.3$), 2.29 (1 H, m), 1.1–2.1 (8 H, m), 2.03 (3 H, s), 0.91 (3 H, s), 0.67 (3 H, d, $J = 6.0$).

Production of Aldehyde 17 from Diene 2. Compound 2 (330 mg, 0.96 mmol) was treated with 11 mL of saturated H_2IO_6 in ether solution for 2.5 h and the filtered. The ether was removed in vacuo to give, in high yield after HPLC purification (50% EtOAc/isooctane), the aldehyde (181 mg, 0.78 mmol, 81%). The aldehyde (29.2 mg, 0.125 mmol) was acetylated under standard conditions to yield, after μ -Porasil HPLC purification (10% EtOAc/isooctane), 17.8 mg (0.065 mmol, 52%) of pure 17, which showed an $[\alpha]_D^{25} +30.1^\circ$ (c 1.75, CHCl_3). The $^1\text{H NMR}$ spectrum of 17 (220 MHz) was superimposable with that of the enone-acetate produced from spatol (1).

Sodium Periodate Cleavage of 2 To Form the Methyl Ketone 18. NaIO_4 (171 mg, 0.8 mmol) was added over a 0.5-h period to a stirred solution containing 134.3 mg of 2 (0.4 mmol), 10 mL of dioxane, and 2.5 mL of H_2O at 0 $^\circ\text{C}$. The reaction was warmed to room temperature and stirred for 3.75 h, at which time 100 mL of diethyl ether was added. The ether was collected, washed with H_2O (3 \times 30 mL), dried over MgSO_4 , filtered, and removed in vacuo to yield, after preparative silica gel HPLC (16 mm \times 50 cm), 116.1 mg of the methyl ketone 18 (0.382 mmol, 96%) as an oil with the following spectral features: $[\alpha]_D^{25} +6.35^\circ$ (c 3.01, CHCl_3); IR (CHCl_3) 3500, 2950, 1675, 1430, 1350, 1200, 1040, 910 cm^{-1} ; $^1\text{H NMR}$ (220 MHz, CDCl_3) δ 6.67 (1 H, dd, $J = 6, 16$), 6.26 (1 H, d, $J = 16$), 5.39 (1 H, s), 5.03 (1 H, s), 4.62 (1 H, d, $J = 6$), 3.75 (1 H, d, $J = 4$), 2.94 (1 H, ddd, $J = 6, 6, 13$), 2.27 (3 H, s), 2.27 (1 H, ddd, $J = 13, 13, 4$), 1.0–2.1 (9 H, m), 0.97 (3 H, s), 0.87 (3 H, d, $J = 6$).

Ozonolysis of 12 To Form the Carbomethoxy Ester 13. The triacetate 12 (258 mg, 0.56 mmol) was treated with O_3 in EtOAc (30 mL) for 30 min, at which time the solution was light purple. Excess O_3 was removed by bubbling Ar through the solution, and 5% Pd/C and H_2 were added under pipet bulb pressure at 0 $^\circ\text{C}$. After 3.5 h the solution was filtered and solvent removed in vacuo. The complex mixture was dissolved in 1 mL of MeOH, and excess ethereal CH_2N_2 was added and allowed to react for 1 h. Removal of excess reactants and solvents in vacuo was followed by chromatography over a short silica column. Fractions eluting with 50% CH_2Cl_2 /isooctane contained 13, which was further purified by using μ -Porasil HPLC (10% EtOAc/isooctane) to give 34.4 mg of 13 (0.123 mmol, 22%) with the following spectral features: $[\alpha]_D^{25} +2.7^\circ$ (c 0.35, CHCl_3); IR (CCl_4) 3000, 1725, 1430, 1360, 1235, 1170, 1025, 965, 890 cm^{-1} ; LRMS (70 eV), m/z (relative intensity) 280 (obsd M^+ , 0.12), 249 (2), 238 (6), 220 (18), 205 (7), 188 (9), 161 (83), 119 (37), 105 (84), 81 (53), 43 (100).

Hydrolysis of Ester 13 To Yield Alcohol 14. The ester 13 (26.5 mg, 0.095 mmol) was stirred for 21.5 h at room temperature in 5 mL of MeOH and 2 mL of saturated Na_2CO_3 , after which the reaction was diluted with 50 mL of H_2O and extracted (3 \times 50 mL) with diethyl ether. The combined ether extracts were dried over MgSO_4 , filtered, and reduced to yield a 1:1 mixture of starting material and desired alcohol by TLC. Preparative

thin-layer chromatography gave a band of pure 14 (10.3 mg, 0.043 mmol, estimated yield 91%) with the following spectral features: IR (CHCl_3) 3500, 2950, 1715, 1430, 1350, 1205, 1100, 1040, 930 cm^{-1} ; $^1\text{H NMR}$ (220 MHz, CDCl_3) δ 3.84 (1 H, d, $J = 4$), 3.68 (3 H, s), 3.28 (1 H, ddd, $J = 6.5, 6.5, 13$), 2.53 (1 H, ddd, $J = 13, 13, 4$), 2.26 (1 H, dd, $J = 7, 7$), 1.1–2.18 (9 H, m), 1.00 (3 H, s), 0.91 (3 H, d, $J = 6$); LRMS (70 eV), m/z (relative intensity) 238 (obsd M^+ , 5), 220 (5), 206 (5), 191 (10), 178 (22), 161 (41), 160 (34), 105 (56), 91 (56), 81 (88), 55 (100).

$^1\text{H NMR}$ Analysis of Alcohol 14 with Incremental Eu(fod)₃ Addition. The incremental addition of Eu(fod)_3 to 14 in CDCl_3 coupled with appropriate spin-decoupling experiments allowed for visualization of the couplings (in hertz) between all protons except those at C-2: $J_{1\beta-11} = 6.0$, $J_{1\beta-9} = 7.0$, $J_{3\beta-10} = 7.0$, $J_{3\alpha-10} = 0$, $J_{9-10} = 7.0$, $J_{9-8} = 6.0$, $J_{8-7\alpha} = 5.0$, $J_{7\alpha-6\alpha} = 13.3$, $J_{7\alpha-6\beta} = 5.0$, $J_{6\alpha-6\beta} = 13.3$, $J_{6\alpha-5\beta} = 0$, $J_{6\beta-5\beta} = 4.4$.

Oxidation of the Alcohol 14 to Ketone 15. A subsequent ozonolysis of 2 followed by treatment with CH_2N_2 , following the procedure previously described, yielded an additional 18 mg of the alcohol-ester 14. The alcohol 14 (12 mg, 0.051 mmol) was treated for 3.5 h with excess pyridinium chlorochromate (PCC) in CH_2Cl_2 (2 mL) with NaOAc buffer present, after which the reaction was quenched with the addition of 20 mL of diethyl ether and filtered through a thin-layer of silica gel. Purification of 15 using a μ -Porasil HPLC column (20% EtOAc/isooctane) yielded 10.6 mg (0.045 mmol, 89%) of product which showed: $[\alpha]_D^{25} -226^\circ$ (c 1.06, CHCl_3); IR (CHCl_3) 2950, 1735, 1460, 1440, 1370, 1230, 1175, 1025, 940 cm^{-1} ; $^1\text{H NMR}$ (220 MHz, CDCl_3) δ 3.73 (3 H, s), 3.16 (2 H, m), 2.52 (2 H, m), 2.36 (1 H, dd, $J = 7.5, 7.5$), 1.0–2.0 (6 H, m), 0.93 (3 H, s), 0.90 (3 H, d, $J = 6$).

Synthesis of Ketone 19 from Alcohol 3. Eight drops of acetic anhydride were added to 46.5 mg of 3 (0.123 mmol) in 1 mL of CH_2Cl_2 and 0.5 mL of pyridine at 0 $^\circ\text{C}$, and the mixture was stirred for 3 h. The solvents and excess reactants were next removed in vacuo. Preparative thin-layer chromatography of the resulting mixture gave a band (30.6 mg, 0.073 mmol, 59%) which contained a mixture of diacetates 21 and 22 and two other bands corresponding to starting material (13.8 mg, 0.037 mmol, 30%) and triacetate 12 (5.3 mg, 0.012 mmol, 9.3%). The band containing the mixture of diacetates was treated for 1 h at 0 $^\circ\text{C}$ in 2 mL of CH_2Cl_2 with excess PCC with powdered NaOAc buffer present. The reaction was quenched with 20 mL of diethyl ether and filtered through a thin layer of silica gel. Thin layer chromatography gave a band containing the two isomeric keto diacetates (19 mg, 0.0455 mmol, 62%) which were partially separated (>90% purity by $^1\text{H NMR}$) by recycling through a μ -Porasil HPLC column (30% EtOAc/isooctane). The desired 5-ring ketone 19 (8.6 mg, 0.021 mmol, 44%) and the cross-conjugated keto diacetate 23 (9.2 mg, 0.022 mmol, 48%) were recovered. Compound 19 showed $[\alpha]_D^{25} -51.5^\circ$ (c 0.67, CHCl_3) and was identical by 220-MHz $^1\text{H NMR}$ with the diacetate produced from 4.

Li(*t*-BuO)₃AlH Reduction of Ketone 4 To Yield 24. Ketone 4 (10.8 mg, 0.0327 mmol) was treated with excess $\text{Li}(t\text{-BuO})_3\text{AlH}$ in 2 mL of absolute EtOH at 0 $^\circ\text{C}$ for 1.5 h. The reaction was quenched with H_2O , and then the ethanol and H_2O were removed in vacuo. The residue was partitioned between H_2O and Et_2O , and the latter was collected, dried over anhydrous MgSO_4 , filtered, and reduced in vacuo. The resultant mixture was treated with acetic anhydride and pyridine (1 mL each) for 18 h at which time excess reactants were removed in vacuo. Preparative thin-layer chromatography yielded a band of pure 24 (8.9 mg, 0.0193 mmol, 59%) which showed the following: $[\alpha]_D^{25} -8.2^\circ$ (c 0.89, CHCl_3); $^1\text{H NMR}$ (220 MHz, CDCl_3) δ 5.85 (1 H, d, $J = 15$), 5.69 (1 H, dd, $J = 7, 15$), 5.53 (1 H, d, $J = 7$), 5.28 (1 H, s), 5.01 (1 H, s), 4.80 (1 H, dd, $J = 10, 8$), 4.14 (1 H, d, $J = 10$), 4.01 (1 H, d, $J = 10$), 2.7–1.45 (11 H, m), 2.12 (3 H, s), 2.09 (6 H, s), 1.34 (3 H, s), 1.02 (3 H, s), 0.92 (3 H, d, $J = 7$).

Reduction and Periodate Cleavage of 5 To Yield the Methyl Ketone 18. Ketone 5 (117.8 mg, 0.312 mmol) was treated with excess LiAlH_4 in tetrahydrofuran (10 mL) for 2.0 h at room temperature and then quenched with ice and H_2O , extracted with ethyl acetate (150 mL), and dried over anhydrous MgSO_4 to give a high yield of tetraol 2 by TLC. This unpurified reaction product was treated with 154 mg of NaIO_4 (0.72 mmol, 2 equiv) in dioxane (10 mL) and H_2O (2.5 mL) for 2 h. The solution was extracted with diethyl ether (3 \times 25 mL), which in turn was washed with

H₂O, dried over anhydrous MgSO₄, filtered, and reduced in vacuo to yield two UV-absorbing products by TLC. Preparative thin-layer chromatography followed by μ -Porasil HPLC (40% EtOAc/isooctane) yielded a 2:1 mixture of 27 and 18. Repeated μ -Porasil chromatography (30% EtOAc/isooctane) eventually separated the two epimers, yielding 18 (7.3 mg, 0.24 mmol, 7.7%) which showed $[\alpha]_D +21.1^\circ$ (*c* 0.44, CHCl₃) and 27 (14.6 mg, 0.048 mmol, 15.4%).

Periodate Cleavage of Tetraol 6 To Yield the α,β -Unsaturated Aldehyde 17. Tetraol 6 (17.2 mg, 0.045 mmol) in 4 mL of dioxane and 1 mL of H₂O was treated with 25 mg of NaIO₄ for 3.25 h. The reaction mixture was extracted with diethyl ether (3 \times 30 mL) which was washed with H₂O, dried over anhydrous MgSO₄, filtered, and reduced. The crude reaction product was treated with excess acetic anhydride (1 mL) in pyridine (1 mL) overnight, at which time the excess reagents and solvents were removed in vacuo. Preparative thin-layer chromatography gave a band of nearly pure 17, which was further purified by using μ -Porasil HPLC (5% EtOAc/isooctane; 5.4 mg, 0.0195 mmol, 43%), showed $[\alpha]_D +26.5$ (*c* 0.54, CHCl₃), and had an ¹H NMR (360 MHz, CDCl₃) identical with that of the enone produced from 1 and 2.

Oxidation of Diol 7 to Diketone 29. Excess pyridinium chlorochromate and NaOAc buffer were added at 0 °C to 16.3 mg of 7 (0.0537 mmol) in 2 mL of CH₂Cl₂ and allowed to react for 2.75 h. Dilution with 20 mL of diethyl ether and filtration through silica gel, followed by removal of the solvent in vacuo, gave a low yield of 29 by TLC analysis. HPLC purification (μ -Porasil, 10% EtOAc/isooctane) yielded 2.7 mg of 29 (0.09 mmol, 17%) which showed the following: $[\alpha]_D -190^\circ$ (*c* 0.27, CHCl₃); ¹H NMR (CDCl₃, 220 MHz) δ 6.05 (1 H, s), 5.84 (1 H, s), 5.00 (1 H, s), 4.93 (1 H, s), 2.93 (3 H, m), 2.39 (4 H, m), 1.94 (3 H, s), 1.2-1.9 (8 H, m), 1.00 (3 H, s), 0.94 (3 H, d, *J* = 7).

Oxidation of Diol 8 To Yield the Diketone 29. To 19.4 mg of 8 (0.0638 mmol) in 2 mL of CH₂Cl₂ were added excess PCC and powdered NaOAc at 0 °C, and the mixture was allowed to react for 2.75 h. The reaction was diluted with 20 mL of diethyl ether, filtered through a thin layer of silica gel, and reduced in vacuo to give a small yield of the diketone 29 by TLC. HPLC purification (μ -Porasil, 10% EtOAc/isooctane) gave 3.8 mg of 29 (0.0126 mmol, 20%) which showed $[\alpha]_D -185.0^\circ$ (*c* 0.38, CHCl₃) and had a 220-MHz ¹H NMR spectrum identical with that of 29 produced from 7.

Ozonolysis and Acetylation of 2 To Yield the Ester 13. The tetraol 2 (150 mg, 0.494 mmol) in 20 mL of EtOAc was treated with O₃ for 25 min at -78 °C, during which time the solution turned a light blue color. Excess O₃ was removed by flushing with Ar, and then 5% Pd/C was added at 0 °C under a slight pressure of hydrogen. The hydrogenation reaction was allowed to warm to room temperature and proceed for 6 h, at which time it was terminated and filtered. Chromatography of the crude mixture on a short silica column gave fractions (50% EtOAc/CH₂Cl₂) containing the alcohol-acid. Treatment of these combined fractions first with CH₂N₂ in Et₂O with 1 mL of MeOH present and then with acetic anhydride in pyridine at room temperature for 14 h provided a crude reaction mixture containing 13. By use of μ -Porasil HPLC (10% EtOAc/isooctane), a low yield of pure 13 was obtained (5.7 mg, 0.020 mmol, 4.1% overall), which showed the following: $[\alpha]_D +1.75^\circ$ (*c* 0.57, CHCl₃); IR (CHCl₃) 1727 cm⁻¹; LRMS (70 eV), *m/z* 280 (obsd M⁺), 249, 238, 221, 220, 205, 188.

Dehydration, Hydrogenation, and Oxidation of 9 To Form Ketone 30. Diol 9 (35.3 mg, 0.116 mmol) was treated with excess *p*-toluenesulfonic acid in aqueous acetone for 1.5 h, at which time the solution was repeatedly extracted with diethyl ether (4 \times 30 mL). The combined extracts were washed with saturated NaHCO₃, dried over anhydrous MgSO₄, filtered, and reduced in vacuo to yield an oily residue. After thin-layer silica gel chromatographic purification, 21.6 mg of the desired dehydration product was recovered (0.076 mmol, 66%). The triene was treated with H₂ under pipet-bulb pressure in the presence of Pt catalyst for 42 h to yield 21.3 mg (0.075 mmol, 99%) of the saturated alcohol. The 60-MHz ¹H NMR spectrum of the crude reaction product showed no bands at lower field than 4.0 ppm and no methyl singlets in the region of 1.5-2.0 ppm. This crude reaction product

was treated with excess pyridinium chlorochromate in CH₂Cl₂ (2 mL) at 0 °C in the presence of powdered NaOAc for 2 h. The reaction was quenched by the addition of 20 mL of diethyl ether and filtered through a thin layer of silica gel to give a modest yield of the saturated ketone 30 after μ -Porasil HPLC purification (10.5 mg, 0.376 mmol, 50.2%, 32% overall) which showed $[\alpha]_D -101.2^\circ$ (*c* 0.37, CHCl₃).

Dehydration, Hydrogenation, and Oxidation of 10 To Form Ketone 30. Diol 10 (27 mg, 0.089 mmol) was treated with excess *p*-toluenesulfonic acid in aqueous acetone for 2.0 h, at which time the solution was repeatedly extracted with diethyl ether (4 \times 30 mL). The combined extracts were washed with saturated NaHCO₃ (3 \times 50 mL) and then H₂O (2 \times 50 mL), dried over anhydrous MgSO₄, filtered, and reduced in vacuo. The unpurified reaction product was treated with H₂ under pipet-bulb pressure in the presence of Pt catalyst for 70 h to yield, by 60-MHz ¹H NMR analysis, the saturated alcohol intermediate as the major product. Oxidation of this crude residue with excess pyridinium chlorochromate in CH₂Cl₂ (2 mL) at room temperature for 1.5 h in the presence of NaOAc buffer followed by dilution with diethyl ether (20 mL) and filtration through a thin layer of silica gel gave predominately the saturated ketone 30 by TLC. Purification of this ketone as produced from 10 by using μ -Porasil HPLC (10% EtOAc/isooctane) gave 10.8 mg (0.039 mmol, 44% overall) of product which showed $[\alpha]_D -99.5^\circ$ (*c* 1.08, CHCl₃).

Hydrogenation and Oxidation of 11 to the Ketone 30. Alcohol 11 (23.1 mg, 0.08 mmol) in 5 mL of diethyl ether was treated with a catalytic amount of Adams catalyst (40 mesh, Alfa) and hydrogen (pipet-bulb pressure) at room temperature with stirring for 40 h, at which time the solvent was filtered and removed in vacuo. The 60-MHz NMR spectrum indicated complete hydrogenation had occurred at this time. This crude reaction product was dissolved in 2 mL of CH₂Cl₂ and treated for 1.25 h at room temperature with excess PCC with powdered NaOAc buffer present. The reaction was terminated with the addition of 20 mL of diethyl ether and filtration through a thin layer of silica gel followed by removal of solvents in vacuo. Purification of the major component by using μ -Porasil HPLC (15% EtOAc/isooctane) yielded 15.0 mg (0.050 mmol, 62.5%) of pure 30 which showed $[\alpha]_D -110.0^\circ$ (*c* 1.50, CHCl₃) and the following ¹H NMR features (360 MHz, CDCl₃): δ 2.53 (2 H, m), 2.28 (3 H, m), 1.0-2.0 (14 H, m), 0.81-1.00 (15 H, m).

Oxidation of Alcohol 11 To Yield Diketone 29. Alcohol 11 (50.1 mg, 0.174 mmol) was dissolved in 10 mL of CH₂Cl₂ in a Pyrex glass tube containing ca. 1.0 mg of methylene blue sensitizer. This solution was irradiated with a 100-W tungsten bulb for 9.5 h, during which time a continuous stream of O₂ was bubbled through the solution. Analysis of the crude reaction product by TLC indicated several discrete UV-absorbing compounds had been produced. HPLC with a μ -Porasil column successfully purified 5(*R*)-hydroxy-17-oxospata-13,18-diene (32; 2.8 mg, 0.0093 mmol, 5.3%). Oxidation of this product in 1 mL of CH₂Cl₂ with pyridinium chlorochromate and powdered NaOAc buffer for 1.75 h led to the exclusive production, after workup by dilution with diethyl ether and filtration through a thin layer of silica gel, of the diketone 29. Final purification involving μ -Porasil HPLC (10% EtOAc/isooctane) yielded 1.0 mg of 29 (0.0027 mmol, 1.6% overall) which showed $[\alpha]_D -150.0^\circ$ (*c* 0.08, CHCl₃) and had a ¹H NMR (360 MHz) spectrum identical with that of the diketone formed from 7 or 8.

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Registry No. 1, 76520-52-0; 2, 77136-60-8; 3, 77136-61-9; 4/5, 77136-62-0; 6, 77136-63-1; (17S)-7/8, 77136-64-2; (17R)-7/8, 77209-25-7; 9, 77136-83-5; 10, 77136-65-3; 11, 77136-66-4; 12/24, 77136-67-5; 13, 77136-68-6; 14, 77136-69-7; 15, 77136-70-0; 16, 77136-71-1; 17, 77136-72-2; 18/27, 77136-73-3; 19/25, 77136-74-4; 20/26, 77136-75-5; 21, 77136-76-6; 22, 77136-77-7; 23, 77136-78-8; 28, 77136-79-9; 29, 77136-80-2; 30, 77136-81-3; 32, 77136-82-4.